

Review

Statins: the new aspirin?

N. R. Veillard and F. Mach *

Cardiology Division, Department of Medicine, University Hospital Geneva, Foundation for Medical Research,
64 Avenue Roseraie, 1211 Geneva 4 (Switzerland), Fax + 41 22 382 7245, e-mail: Francois.Mach@medecine.unige.ch

Received 6 June 2002; received after revision 6 September 2002; accepted 6 September 2002

Abstract. 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors, or statins, have been described as the principal and the most effective class of drug to reduce serum cholesterol levels. Statin therapies have been shown to reduce cardiovascular events, including myocardial infarction, stroke, and death, significantly, by altering vascular atherosclerosis development in patients with or without coronary artery disease symptoms. Extensive use of statins has led to the increase of

some undesirable effects that are heavily counterbalanced by the benefits. Indeed, pleiotropic effects extend far beyond cholesterol reduction and involve non-lipid-related mechanisms that modify endothelial functions, immunoinflammatory responses, smooth muscle cell activation, proliferation and migration, atherosclerotic plaque stability, and thrombus formation. In this review, we describe in detail the targets and mechanisms of action of statins.

Key words. Statin; atherosclerosis; inflammation; cholesterol; pleiotropic effect.

Introduction

Atherosclerosis is a chronic immunoinflammatory disease, which very often begins in early childhood and can lead to severe clinical manifestations later in life. Atherosclerotic vascular lesions are characterized by accumulation of lipids, fibrous elements, and immune cell infiltrates. Weakening of the fibrous cap covering an atherosclerotic lesion causes plaque rupture and thrombosis, crucial features inducing acute clinical complications such as unstable angina (UA), myocardial infarction (MI), or cerebral stroke [1–3]. Over the past few decades, atherosclerosis has been established as the most common cause of death in Western countries, responsible for half of the morbidity and mortality. Every year, 1.5 million over 250 million Americans are hospitalized for acute coronary syndromes, either UA (50%) or MI (50%). After acute MI, patients remain at high risk for recurrent coronary events and cardiovascular mortality. Indeed,

within one year after an acute MI, 25% of men and 38% of women will die from vascular diseases [4, 5], and MI will reoccur in 18% of men and 35% of women within 6 years after a clinical event. Cerebral stroke leads to serious disabilities: among stroke survivors, 50% have hemiparesis, 33% are clinically depressed, 25% are unable to walk, and 17% are aphasic [6]. Silent strokes or infarct-like lesions (ILLs) increase with age with an incidence of 22% in patients over 65 years compared to 43% in patients older than 80 years [7].

Cardiovascular risk factors are numerous and vary with age. The increased incidence of cardiovascular events in Western countries mainly results from hypercholesterolemia, hypertension, smoking, diabetes, obesity, genetic backgrounds, and prolonged life span. Recent results have shown that mental stress may also participate in the development of atherogenesis [8, 9]. 3-Hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors, or statins, have been described as the principal and most effective class of drugs to reduce serum cholesterol levels, and have been shown to reduce significantly

* Corresponding author.

cardiovascular events and mortality in patients with or without coronary artery disease. Due to their broad and extensive clinical beneficial effects, some of which are unrelated to lipid-lowering effects (see below), and their extremely low incidence of side effects, statins could be perceived as the new aspirin.

Structure and properties of statins

Looking to the microbial world, researchers were hoping to find a microorganism that produced an HMG-CoA reductase inhibitor as a defense mechanism against attack by other microbes which relied on sterols as part of their biochemical make-up [10]. *Pythium ultimum* was found to produce an anti-fungal substance called citrinin [11], which irreversibly inhibits HMG-CoA reductase. A new compound known as mevastatin was isolated from a second mold, *Penicillium citrinum* [12], and investigations showed that it was capable of inhibiting lipid synthesis. By 1976, Endo and Kuroda had isolated a similar molecule, lovastatin, from *Aspergillus terreus* [12]. The new compound was slightly more effective as an HMG-CoA reductase inhibitor than mevastatin. Further development of drugs based on mevastatin and lovastatin has followed three main approaches. First, synthetic compounds, such as fluvastatin, were produced by replacing the decalin ring of the fungal compounds with an aromatic ring. Second, chemical alteration of fungal products created drugs such as simvastatin. Finally, microbial alteration of fungal compounds has led to drugs such as pravastatin. The affinity of HMG-CoA reductase is substantially higher for the natural statins (in the case of mevastatin, 10,000 times higher) than it is for HMG-CoA; mevastatin acts as a reversible competitive inhibitor to the enzyme reaction, and less mevalonic acid is produced in its presence. On the other hand, synthetic statins model the product mevalonate. Thus, the cholesterol production pathway is broken. The introduction of a competitive inhibitor for HMG-CoA reductase results in two physiological responses. In compensation for the inhibition, cells begin to produce more HMG-CoA. The direct reduction in circulating cholesterol is therefore only small. However, the number of low-density lipoprotein (LDL) receptors on the cell surface of patocytes increases markedly [13]. As the liver is responsible for removing LDL cholesterol from plasma via the LDL receptor mechanism, blood cholesterol levels also fall dramatically.

Because of the increased incidence of cardiovascular events in western countries, statins have interested the pharmacological industry. Since the development of lovastatin and its approval in 1987, 11 other statins have been developed or are in the process of being studied. Five of these are actually commercialized: atorvastatin, fluvastatin, lovastatin, pravastatin, and simvastatin. Ator-

vastatin, pravastatin, and simvastatin represent almost 70% of statins used in Europe and the United States [14]. Two subtypes of statins are available on the market: the natural (fermentation-derived) and the synthetic statins (fig. 1). Molecules derived by fermentation, including lovastatin, pravastatin, and simvastatin have very similar chemical structures. Simvastatin is approximately twice as potent as pravastatin and lovastatin, whilst mevastatin is the least powerful. On the other hand, the structures of the synthetic statins, atorvastatin, cerivastatin, and fluvastatin, are very different. The described structural characteristics are closely related to the physicochemical properties of HMG-CoA reductase inhibitors. By altering the basic chemical composition of the mevastatin molecule, drug potency can be increased. Statins differ in their lipophilicity/hydrophilicity (table 1) [15], which reflects their potential to cross cellular membranes nonselectively by passive diffusion, and explain why pravastatin does not easily cross cellular membranes whereas lovastatin and simvastatin do [16]. Large clinical human studies have demonstrated that statins reduce total serum cholesterol by 15–40%, LDL cholesterol by 20–60%, triglycerides by 10–30%, and increase high-density lipoprotein (HDL) cholesterol by 5–15% [17]. LDL plasma levels are more effectively reduced by the latest developed atorvastatin compared to the older statins [18], with an incremental reduction in LDL by 6–7% when the statin dose is doubled [19]. Nevertheless, compared to other statins, atorvastatin does not increase HDL cholesterol in a dose-dependent manner [20].

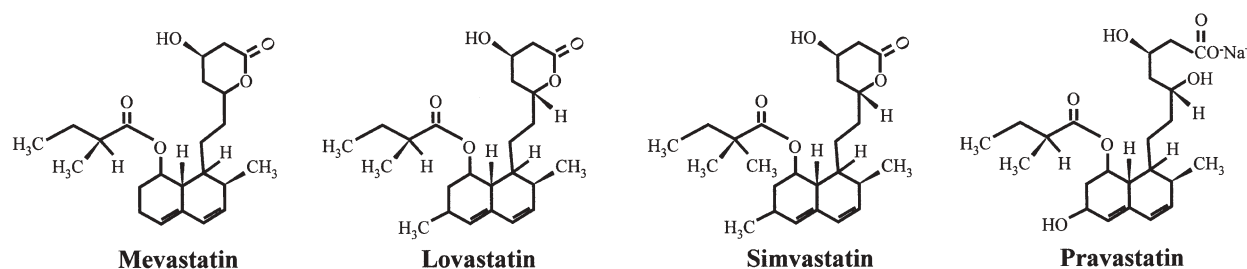
Mechanism of action

By inhibiting L-mevalonic acid synthesis, statins also prevent the catabolism of other isoprenoid intermediates of the cholesterol biosynthetic pathway (fig. 2), such as farnesylpyrophosphate (FPP) and geranylgeranylpyrophosphate (GGPP) [21]. Thus, since mevalonate is not the only precursor of cholesterol, statins have pleiotropic effects by concomitant regulation of numerous other mevalonate metabolites [21–24]. Indeed, isoprenoids are vital for multiple cellular functions, such as covalent attachment, and GGPP and FPP prenylation of numerous

Table 1. Comparison of the lipophilicity of statins.

Statins	Lipophilicity at pH 7.4 (log + SE)
Cerivastatin	1.69 + 0.02
Simvastatin	1.60 + 0.06
Fluvastatin	1.27 + 0.07
Atorvastatin	1.11 + 0.02
Rosuvastatin	−0.33 + 0.06
Pravastatin	−0.84 + 0.06

Fermentation-Derived Statins



Synthetic Statins

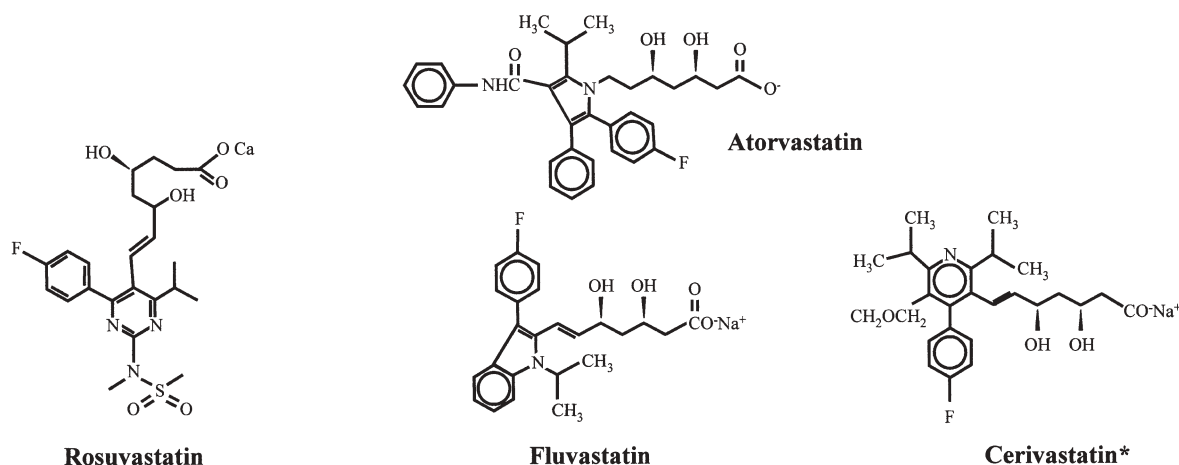


Figure 1. Chemical structures of fermentation-derived (natural) statins and synthetic statins. *Cerivastatin was removed from the market since August 2001.

proteins. Prenylation of proteins is a prerequisite for the cell membrane association of both plasma and internal membranes, and is essential for their functions [25, 26]. These prenylated proteins include the γ subunit of heterotrimeric G proteins, heme-a, nuclear lamins, and the GTP-binding proteins Ras and Ras-like proteins (Rho, Rab, Rac, Ral, and Rap) [27]. Within endothelial cells, the posttranslational modification of Ras is FPP dependent, whereas Rho is GGPP dependent [28, 29]. Proteins of the Rho family are implicated in cell shape, cytoskeleton organization, motility, secretion, proliferation, and cell signaling. In response to extracellular signals such as growth factors or during cellular proliferation, migration or mitosis, cells undergo reorganization of their actin cytoskeleton, which alters the three-dimensional colocalization of their proteins [27–30]. Thus, Rho-induced changes in the actin cytoskeleton may affect intracellular transport, membrane trafficking, mRNA stability, and gene transcription [31]. Statin inhibition of the Rho kinase pathway, a downstream Rho target, leads to the accumulation of inactivated Rho within the cell cytoplasm, which is believed to have pleiotropic effects on vascular cells, leukocytes, and bone [29, 32–35]. In addition, sev-

eral studies have shown that statins decrease the expression and secretion of many different immunoinflammatory molecules, which could be prevented by the addition of L-mevalonate and, in some cases, by farnesol or geranylgeraniol [23, 29, 36–39].

By inhibiting the RhoA signal transduction pathway [40], statins are able to activate peroxisome proliferator-activated receptor (PPAR) α , β/δ , and γ in a dose-dependent manner [41], an effect which could be reversed by FPP, GPP, cholesterol, or sterol regulatory element-binding protein-1 (SREBP-1) [41]. PPARs constitute a subfamily of the nuclear receptor family [42] comprising three isotypes, PPAR α , β , and γ , which are characterized by distinct tissue [43, 44] and developmental distribution patterns [45]. PPARs are ligand-activated transcription factors which, upon heterodimerization with the retinoic X receptor (RXR), bind to specific peroxisome proliferator response elements (PPREs) located in the promoter of target genes [46]. PPARs exhibit direct anti-inflammatory properties beside their crucial role in β -oxidation of fatty acids and arachidonic acid metabolites [47]. Indeed, PPARs are involved in the control of vascular inflammation and thrombogenicity related to atherosclerosis [48,

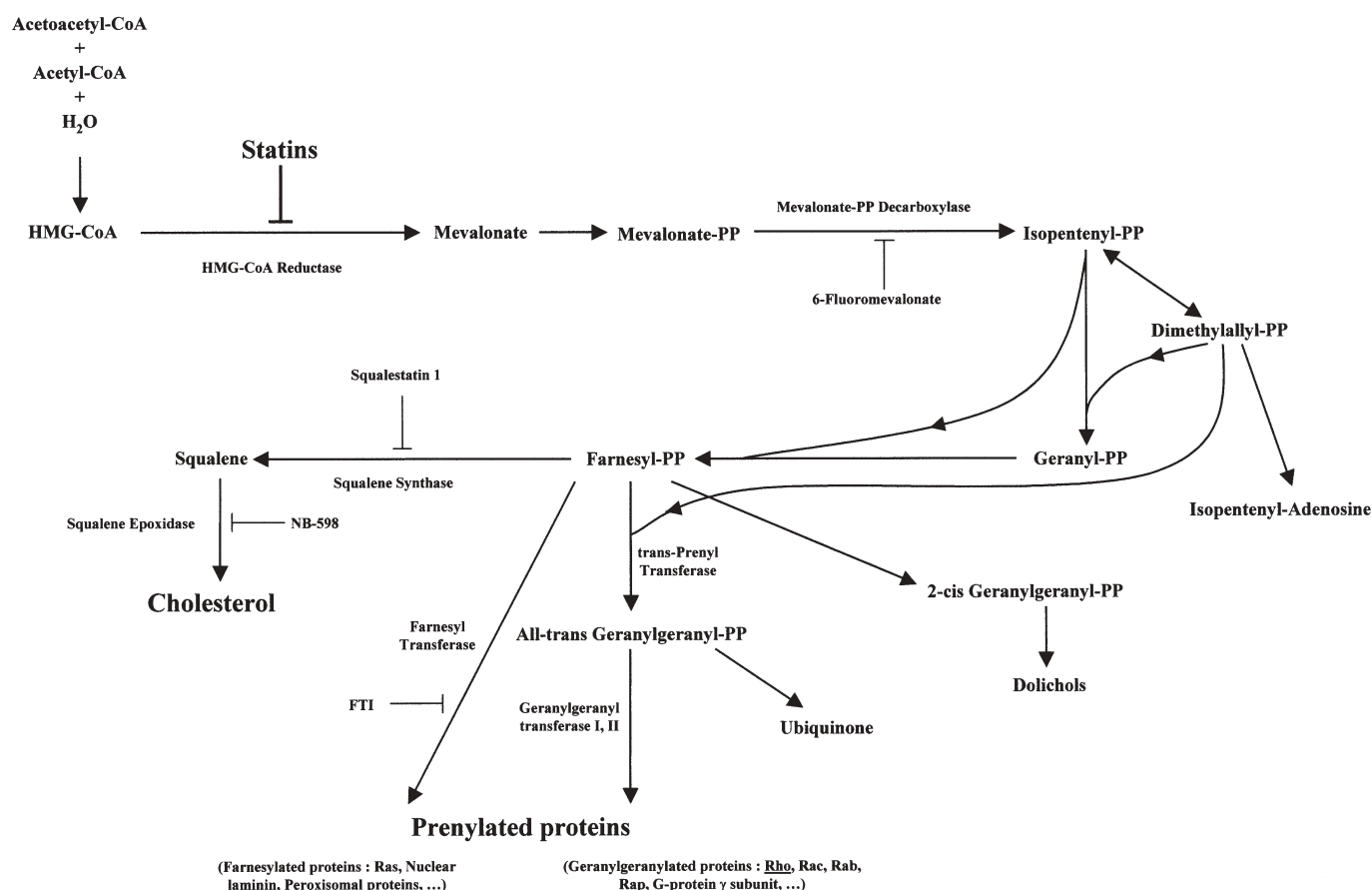


Figure 2. The metabolic pathway of mevalonate and cholesterol. The enzymatic steps and their inhibitors are indicated.

49]. PPAR α has been shown to exert anti-inflammatory actions by preventing the binding of the activator protein-1 (AP-1) and the nuclear factor kappa B (NF- κ B) protein to their DNA target sequences [50–52]. The activation of NF- κ B and the synthesis of numerous NF- κ B-dependent gene products have been linked to the activation of inflammatory cells [53] (fig. 3). Recent reports have shown that NF- κ B was activated in 60% of atherosclerotic lesions of untreated mice, especially within macrophages and smooth muscle cells (SMCs), compared to only 30% for atorvastatin-treated animals [54]. In addition, SMCs activated with tumor necrosis factor alpha (TNF- α) and treated with atorvastatin showed reduced activity for NF- κ B and decreased expression of the chemokine MCP-1 [54], an important chemoattractant protein crucial for the recruitment of inflammatory cells [55]. Of note is that NF- κ B is one of the most pleiotropic transcription factors, and is the main nuclear factor involved in the activation of MCP-1 transcription [56, 57]. Thus, mechanisms of anti-inflammatory properties of statins may be in part mediated via PPARs (fig. 3).

Pharmacokinetic effects

Absorption

With the exception of lovastatin and simvastatin, which are administered as lactone prodrugs and must be hydrolyzed in vivo to the corresponding β -hydroxy acid to achieve pharmacologic activity, all statins are administered as the active β -hydroxy acid form.

The extent of absorption of statins varies from 30 to 98% [15, 58, 59]. All statins are absorbed rapidly following oral administration, with time to peak concentration reached within 4 h. Fluvastatin is the only statin able to achieve peak plasma concentration in the micromolar range, which may explain its direct antiatherosclerotic properties. Food intake has variable effects on the absorption of statins, with decreased bioavailability for fluvastatin, pravastatin, and atorvastatin. However, lipid-lowering effects of statins do not differ when taken with the evening meal or at bedtime. The plasma half-life is 2–3 h for all statins except atorvastatin, which has a half-life of 14–20 h [18]. Recently, fluvastatin has also become available in a slow-release form. The pharmacokinetic half-lives of statins do not correspond to the dura-

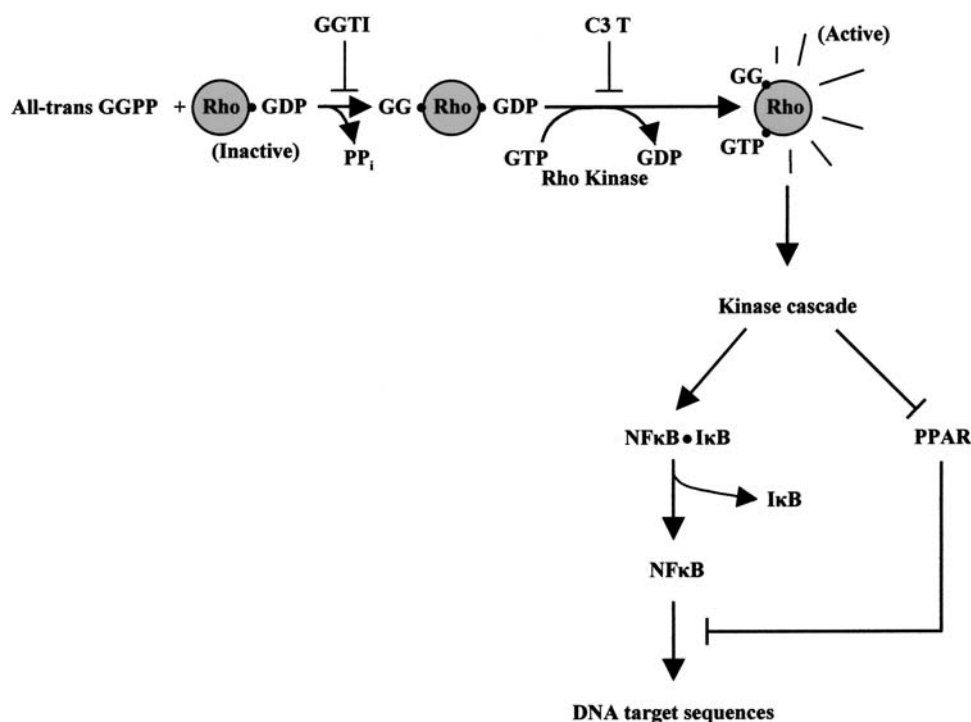


Figure 3. The RhoA prenylation pathway and its main targets.

tion of their pharmacodynamic properties. Due to its long half-life, atorvastatin can accumulate in plasma, achieving a steady-state drug concentration after multiple doses.

Distribution

Except for pravastatin, all statins are highly bound to plasma proteins (mainly albumin). Therefore, systemic exposure to unbound active drugs remains extremely low. In addition, since statins are highly extracted by the liver, drug displacement interactions are also limited. As a result of low plasma protein binding, unbound pravastatin is tenfold higher than the unbound form of other statins [60]. Nevertheless, widespread tissue distribution is prevented by the high hydrophilic properties of pravastatin. With the exception of pravastatin, all statins and their metabolites are excreted via the bile into feces. The amount of statin excreted in urine is negligible, except for pravastatin, which can reach 20% [61, 62].

Metabolism

Almost all statins are metabolized by cytochrome P450 (CYP450) enzymes (3A4, 2C9, 2D6, 1A2) [63] present in the liver, but also in the intestine. Numerous drugs that can be administered concomitantly with statins are often inhibitors of CYP 3A4 (e.g., cyclosporine A, verapamil, diltiazem, and grapefruit juice), and therefore might re-

duce or block the metabolism of certain statins. This inhibition of statin metabolism can raise their plasma concentration above a toxic threshold, leading to muscular lysis. Furthermore, some statins are transported to the intestinal lumen via P-glycoprotein (P-gp) [59], and most CYP 3A4 inhibitors are also P-gp inhibitors. Thus, these drug interactions often result in the coinhibition of their hepatic and intestinal CYP 3A4-dependent metabolism and of their P-gp-dependent transport [64]. For example, coadministration of a statin and cyclosporine A increases the incidence of toxicity. Among statin molecules, some can interact with other medications, depending on their hepatic and intestinal metabolism. In contrast, certain statins do not, or interact less through the CYP 3A4-dependent metabolic pathway [64]. Pravastatin has multiple metabolic pathways, thus decreasing the likelihood of any drug interactions with the CYP 3A4 enzyme. The statin fluvastatin is predominantly metabolized by the cytochrome CYP 2C9, and also by CYP 3A4 and CYP 2C8, and the use of CYP 2C9 inhibitors is rare [65]. Cerivastatin and atorvastatin are metabolized in part by CYP 3A4, but if this cytochrome is inhibited they will be eliminated via other metabolic pathways with a decreased degradation rate. The statins simvastatin and lovastatin are almost exclusively CYP 3A4 dependent. Although pharmacokinetic effects are of great importance, most of the drugs that might cause interaction problems, like erythromycin and cyclosporine, can be managed clinically. Indeed, low doses of statin used with cyclosporine A

seem to be very safe, and statin therapy can easily be interrupted for some days if needed.

Toxicity

The benefits of HMG-CoA reductase inhibitors are supported by extensive literature. However, the growing interest in statins and concomitant increase in prescription rates have led to a rise in undesirable side effects such as hepatotoxicity and myopathy. In August 2001, cerivastatin was reported to cause severe myopathy and rhabdomyolysis with an unexpectedly high frequency compared to other statins of equal or greater efficacy [66, 67]. Cerivastatin has since been removed from the market. These clinical symptoms occur in 1 out of 1000 patients, however, if the clinical events related to cerivastatin are excluded, this ratio is reduced to roughly 1 out of 10,000 patients for all statins together [14]. In changing the shape of the active molecule, the chances and severity of side effects are also altered. For example, there is an increased risk of muscle toxicity with lovastatin in comparison to pravastatin. Although the incidence seems to be dose-dependent, the explanation of myotoxicity is still unclear and controversial, and many hypotheses have been proposed.

Most of the HMG-CoA reductase inhibitors are metabolized by the liver. Damage to this organ is assessed by a persistent elevated level of its transaminases: aspartate transferase (ATS) and alanine transferase (ALT) [14]. Statin treatment commonly leads to a small increase of ALT. Levels of ALT more than three times the upper normal limit indicate a potential liver toxicity (cholestatic jaundice) in patients. Up to 5% of patients will show a slight increase in ATS or ALT at about 6 weeks after initiation of therapy [14]. Transaminase elevation up to the toxicity threshold is present in 1–2% of patients under statin treatment, and clinical symptoms are dependent on patients [68, 69]. In addition, the frequency of persistent transaminase elevation is consistent with all commercialized statins, and is dose dependent [70].

Pharmacodynamics

Although any drug should be administered with appropriate caution, it can be said that the clinical benefits of statin therapy far exceed the very low incidence of risk. Indeed, the beneficial effects of statins extend far beyond their lipid-lowering properties. The LDL cholesterol level is an important target for cardiovascular prevention and the concept of LDL plasma cholesterol level magnitude has been tightly correlated with arteriosclerosis lesion extent and clinical cardiovascular events. However, only half of individuals suffering from an elevated LDL plasma cholesterol level will die from coronary heart dis-

ease (CHD) [71, 72]. Serum total cholesterol and CHD have been correlated [72–74]; however, to improve and better define CHD risk, measurements of HDL [75], fibrinogen [76, 77], plasma viscosity [78] and C-reactive protein (CRP) [76] are very helpful. Several clinical studies have demonstrated that statin therapy reduces CRP levels independently of lipid-lowering effects (see below).

HMG-CoA reductase inhibitors may delay or improve the pathogenesis and clinical symptoms of Alzheimer's disease (AD) [79, 80]. Recent studies suggest that lipids play an important role in the development of AD [80]. Indeed, apolipoprotein (ApoE) $\epsilon 4$, an allele of Apo E, is the major genetic risk factor for AD [81]. Furthermore, the isoform ApoE $\epsilon 4$ correlates with an increased risk for atherosclerosis [82] and amyloid plaque formation [83]: a high cholesterol plasma concentration increases amyloid plaque formation [84, 85]. Since the cellular cholesterol level affects neuronal beta-amyloid peptide (A β) production in vitro [86], and A β has been found to be the major constituent of amyloid plaque, the serum cholesterol concentration might be correlated with A β protein expression. In retrospective studies, statin therapy has shown to reduce the occurrence of AD by 70% [87]. However, AD may also occur because of other biological factors, as is the case with atherosclerosis. Recent in vitro and in vivo experiments have highlighted the antiatherosclerotic effects of statins by lipid-independent mechanisms. These pleiotropic effects of statins on the development of atherosclerosis lesions are described below.

Improvement of endothelial function

Nitric oxide (NO) synthesis is a crucial mediator of vascular homeostasis and blood flow. Decrease of NO synthesis by vascular endothelial cells promotes, in part, vasoconstriction, platelet aggregation, and leukocyte recruitment and adhesion [88–93]. Knock-out mice lacking endothelial NO synthase (eNOS) revealed increased arterial blood pressure and exhibited larger cerebral infarctions after middle cerebral artery occlusion [92–94]. Moreover, cerebral blood flow is reduced and postfocal ischemia tissue damage is induced when eNOS activity is inhibited [95]. In contrast, enhanced NO production by administration of either an NO donor or the eNOS substrate, L-arginine, confers stroke protection after induction of cerebral ischemia [96–98]. Statins can directly upregulate eNOS expression in vitro under cholesterol-clamped condition [29, 99, 100]. Indeed, beneficial effects of statins are absent in eNOS-deficient mice [101, 102]. Statins increase eNOS expression by extending eNOS mRNA half-life, but not eNOS gene transcription [28]. Furthermore, statins reduce in vivo cerebral infarct size [101] as well as oxidative stress [103], and improve neurological function in normocholesterolemic mice

[101]. However, statins have additional effects on endothelial cells. Expression of the procoagulant tissue factor induced by thrombin is prevented by simvastatin treatment through inhibition of Rho/Rho kinase and activation of Akt [104]. Statins also increase the expression of tissue-type plasminogen activator [37, 105] and inhibit the expression of endothelin-1 [38], a potent vasoconstrictor and mitogenic molecule that regulates vascular tone and remodeling [106]. These findings suggest anti-atherothrombotic effects of statins that might be very relevant in the prevention of acute coronary syndromes [107].

Reduction of inflammation

Endothelial dysfunction, related to vascular injury in response to cardiovascular risk factors, triggers the migration of leukocytes within the vessel wall, mainly that of monocyte/macrophage and T lymphocyte types [108]. Adhesion molecules, proinflammatory cytokines and chemokines mediate the extravasation of inflammatory cells. Within atherosclerotic sites, endothelial cells and leukocytes both increase their expression of numerous adhesion molecules and counter receptors, such as the intercellular adhesion molecule-1 (ICAM-1) [109], vascular cell adhesion molecule (VCAM-1), β_1 -integrin and β_2 -integrin [110], and P-selectin [102]. Blocking these adhesion molecule interactions by administration of antibodies or gene targeting (knock-out) has been found to decrease atherosclerotic lesion formation in vivo, indicating a potential therapeutic role for inhibition of leukocyte adhesion and extravasation [111, 112]. Indeed, several in vitro studies have described the beneficial effects of statins by decreasing adhesion molecules such as the monocytic CD11b [113] and the leukocyte function antigen-1 (LFA-1) [114, 115]. Of note is that the expression LFA-1 is HMG-CoA reductase independent; statins can bind directly via a novel regulatory site of β_2 -integrin, which serves as a major counter receptor for ICAM-1 on leukocytes [114]. In addition, other in vitro studies have demonstrated that statins could also reduce the secretion of the proinflammatory cytokines interleukin (IL)-6 and IL-1 β but not TNF- α , the chemokines IL-8, IP-10 and MCP-1, as well as the important immunoregulator CD40 [F. Mulhaupt and F. Mach, unpublished data; 54, 88, 116–122]. All these observations support recent human studies suggesting that statins reduce the number of inflammatory cells within atherosclerotic plaques [123, 124]. The inhibitory effect of statins on leukocyte recruitment was confirmed by in vitro measurements on monocytic cells (U937) pretreated with cerivastatin. This treatment reduced the adhesion of U937 cells to an activated endothelium by the downregulation of CD11a, CD8, and VLA4 [125], an effect that could be reversed by mevalonate. Furthermore, Yoshida et al. [125] demon-

strated that cerivastatin also reduced F-actin polymerization via RhoA inhibition within monocytes. These results were confirmed by the preincubation of monocytic cells with C3 transferase, an exoenzyme inhibiting Rho by ADP ribosylation, which could reduce monocyte adhesion to endothelial cells [125, 126]. However, the expression level of integrins on the U937 cell surface was not significantly altered. Protein geranylgeranylation, specifically RhoA geranylgeranylation, was recently reported to be required for integrin-dependent adhesion of leukocytes [127]. Therefore, RhoA may modulate the affinity of integrins. Indeed, as mentioned in another report, RhoA is required for the clustering of adhesion molecules, such as E-selectin, ICAM-1, and VCAM-1 on the endothelial cell surface when monocytes adhere to the endothelium [126]. These data suggest that Rho is required in endothelial cells for the assembly of stable adhesions with monocytes via the clustering of monocyte-binding receptors and their association with the actin cytoskeleton [126]. Similar but complementary in vivo experiments [102] with the new statin rosuvastatin (not yet commercially available) showed important anti-inflammatory effects via inhibition of endothelial cell P-selectin expression, a protective action mediated by vascular endothelial NO. Rosuvastatin had no effect on leukocyte-endothelium interactions in eNOS-deficient mice, thus underlining the crucial anti-inflammatory role of NO. Since P-selectin stored in Weibel-Parade bodies within endothelial cells is translocated to the cell surface upon stimulation by various inflammatory mediators (such as histamine, thrombin, and oxygen-derived free radicals [128]) via a process modulated by NO [129], increased NO production by statins could explain the modulation of these leukocyte-endothelium interactions.

The reduction of inflammation by statin treatment has been demonstrated by the reduction of high-sensitivity CRP (hs-CRP), a clinical marker of inflammation produced by the liver in response to proinflammatory cytokines such as IL-6 [130, 131]. The level of CRP expression is elevated in patients with coronary artery disease, coronary ischemia, and MI compared to normal subjects [132, 133]. Statin therapy lowers CRP levels, without correlation with lipid lowering (either LDL or total cholesterol) [70, 131, 134–136]. Indeed, almost all patients who clinically benefit from statin therapy had abnormal elevated CRP values [137].

SMC proliferation

SMC proliferation is a central event in the pathogenesis of vascular lesions such as atherosclerosis, postangioplasty restenosis, or transplant arteriosclerosis. Stimulation by chemokines or growth factors, released from endothelial cells, macrophages, or T cells, causes activation, proliferation, and migration of SMCs from the media to

the intima [138, 139]. Independently of their lipid-lowering properties, statins (simvastatin, cerivastatin, and fluvastatin) have been reported to reduce in a dose-dependent manner SMC migration and proliferation in vitro [39, 140, 141]. This inhibitory effect was prevented in vitro by addition of mevalonate, all-trans farnesol (F-HO) and all-trans geranylgeraniol (GG-OH), but not by 2-cis GG-OH, squalene, or ubiquinone [140]. In the same direction, statin treatment has demonstrated a beneficial effect on the rate of restenosis as well as reducing major adverse cardiac events following percutaneous transluminal coronary angioplasty (PCI) in human studies [142–144]. Statins may affect cell growth via interference with signaling pathways that require prenylated proteins [21, 26]. Isoprenoid intermediates may also regulate platelet-derived growth factor (PDGF), which is implicated in SMC DNA synthesis [33]. Whereas statins inhibit PDGF, they also downregulate retinoblastoma gene products and cyclin-dependent kinase (cdk)-2, -4, and -6 activity [33]. In contrast, Laufs et al., [33] demonstrated that the cdk inhibitor protein $p27^{Kip1}$, which binds to and inhibits the activation of cdk-cyclin complexes, increased under statin treatment, but no changes were observed for p16 and $p21^{Waf1}$ cdk inhibitors or the p53 tumor suppressor gene [33]. This upregulation of $p27^{Kip1}$ was linked to the inhibition of Rho but not of Ras, due to the reverse effect of GGPP and not of FPP. Furthermore, addition of C3 or N19RhoA could also increase $p27^{Kip1}$ and inhibit retinoblastoma hyperphosphorylation, resulting in the release of the transcription factor E2F, which induces the expression of genes required for progression through S, G2, and M mitosis phases [145]. In addition, activation of Rho GTPase decreased $p27^{Kip1}$ and increased SMC DNA synthesis, suggesting that the downregulation of $p27^{Kip1}$ by Rho GTPase is mediated by PDGF. These findings are consistent with recent studies showing that elimination of $p27^{Kip1}$ during the G1 to S phase is required for the growth activation of FRTL-5 cells, and that GGPP but not FPP restores the inhibitory effect of statins on the degradation of $p27^{Kip1}$ and allows cdk2 activation [146]. Statins have been reported to cause a significant and dose-dependent reduction in cell proliferation (vascular SMCs in culture). This delay of cycling cells in the G1 and G2/M phases, a phenomenon reversible by mevalonate, leads to apoptosis in different SMCs of different origin [36, 147]. Statins (simvastatin, lovastatin, and fluvastatin) not only decrease SMC migration and proliferation in vitro, but also in vivo [36, 148]. Stark and colleagues [149] showed that geranylgeranylated proteins were not only required for growth but also for apoptosis protection. Indeed, recent studies have highlighted the possible implication of apoptosis in SMC proliferation within atherosclerotic lesions [150, 151].

Apoptosis

Programmed cell death of vascular SMCs has recently been identified in physiological remodeling of the vasculature, as well as in atherosclerosis and restenosis. Statins induced apoptosis of endothelial cells and SMCs in a dose-dependent manner [152, 153], an effect that could be reversed by L-mevalonate, FPP, and GPP, but not by isopentenyl adenosine, ubiquinone, or squalene. Prenyl transferase inhibitors confirmed the involvement of prenylated proteins in SMC apoptosis. Treatment with statin (atorvastatin, lovastatin, and simvastatin but not pravastatin) was associated with a decrease in the prenylation of p21-Rho B, a protein that plays an important role in the control of apoptosis [152]. Human vascular SMCs express the receptor Fas, whereas Fas ligand is expressed on endothelial cells, macrophages, and T lymphocytes, indicating that Fas/Fas ligand-mediated apoptosis may occur in vascular cells at sites of high inflammation [154]. Stimulation with the cytokines interferon (IFN)- γ , TNF- α and IL-1 β induces Fas expression and thereby enhances the apoptotic effect of Fas in certain cell types [154, 155]. The elevated secretion of proinflammatory cytokines and chemokines from vascular endothelial cells, SMCs, and inflammatory cells may increase the apoptotic rate of these cells within the neointima of atherosclerotic plaques. Other pathways may induce apoptosis, such as c-myc, p53, and Bcl-2. Apoptosis is also regulated by insulin-like growth factor (IGF-1) and PDGF, which are survival factors reduced by statins in vitro [33, 156]. In contrast, apoptosis of SMCs is decreased in human carotid plaques of pravastatin-treated patients [124]. Statin treatment may reduce apoptosis by decreasing proinflammatory mediators as well as inflammatory cells within atherosclerotic lesions.

Plaque stability and thrombosis

The composition and tensile strength of an atherosclerotic plaque, rather than its size, are the most important determinants of atherosclerosis complications (fig. 4). Advanced atherosclerotic plaques are characterized by a highly thrombogenic lipid core covered by a thin fibrous cap composed of SMCs and extracellular matrix [157]. Collagen, secreted by SMCs, is the main component of the fibrous cap, and is responsible for its tensile strength [157]. In addition to collagen, paucity of vascular SMCs and prominent accumulation of inflammatory cells are the characteristic components of atherosclerotic plaques prone to rupture with further thrombus formation and consequent clinical cardiovascular events [158]. The degradation of collagen plays an important role in the development and subsequent instability of the plaque. Secretion of proteolytic enzymes, such as matrix metalloproteinases (MMPs), by SMCs and macrophages affect the fibrous content of atherosclerotic lesions, particularly

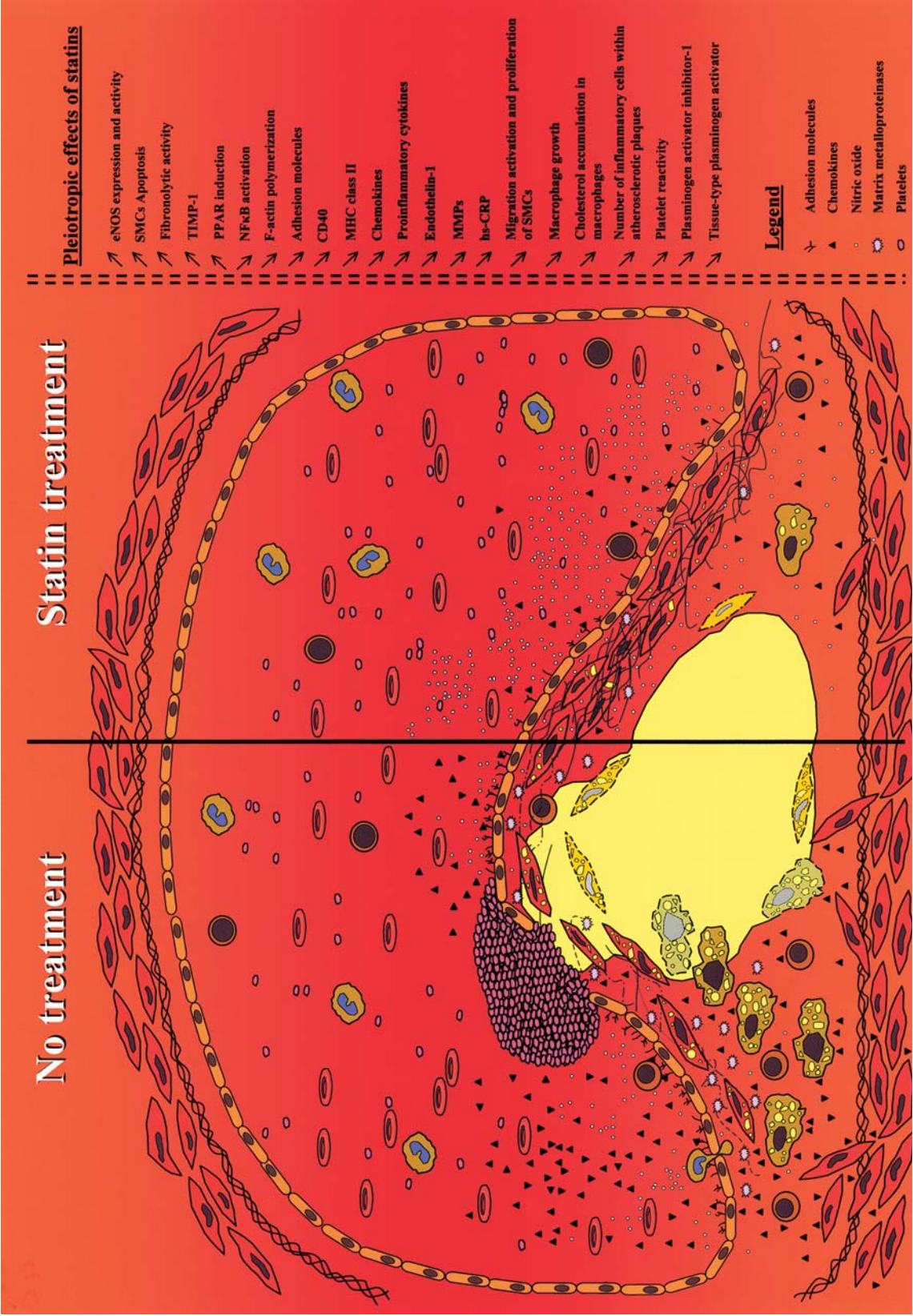


Figure 4. Schematic drawing of an advanced atheroma. Necrosis of macrophages and SMC-derived foam cells leads to the formation of a necrotic core with accumulation of extra-cellular cholesterol. On the left side are represented the major pathophysiological features occurring within an arterial vessel wall of a non-statin treated patient not treated with statins. Numerous inflammatory cells, poor SMCs and collagen content of the fibrous cap, degraded principally by the matrix metalloproteinases often lead to plaque rupture followed by thrombus formation. On the right side are represented the main beneficial effects of statin therapy resulting in a more stable atherosclerotic plaque.

at the shoulder region of the plaque, the margin between the lesion and the unaffected portion of the artery. Indeed, this vulnerable region is predominantly inhabited by foam cell macrophages [158]. In addition, macrophages are capable of weakening the extracellular matrix by phagocytosis [159]. Furthermore, apoptosis of SMCs and macrophages, resulting from cell-cell interactions and the local cytokine environment, involving pro- and antiapoptotic proteins that include death receptors, proto-oncogenes, and tumor suppressor genes, also appears to influence the stability of the plaque [2, 160]. Thus, fissuring, erosion, and ulceration of the fibrous cap leads to plaque rupture followed by thrombosis [161]. Plaque disruption with superimposed thrombosis is the main cause of acute coronary syndromes such as UA, MI, and sudden death [158].

Although changes in plaque size by lipid-lowering treatment may occur over an extended time and are quite minimal [162], statins may contribute to plaque stability mainly by modifying cellular composition and physicochemical properties of atherosclerotic plaques. Statins decrease monocyte/macrophage infiltration as well as the proliferation and migration of SMCs into the neointima. Statins also reduce the secretion of MMPs by SMCs [163] and macrophages [164–166]. MMP-1, MMP-3, and MMP-9 secretion could be inhibited *in vitro* in a dose-dependent manner in macrophages and other vascular cells [164, 167, 168]. Similar results were obtained for MMP-2 in human carotid plaques with prospective statin treatment [124]. The nuclear factor NF- κ B (regulated by statins) has also been demonstrated to control the secretion of MMP-1 and MMP-3, but not the expression of MMP-9, which might be linked to another regulation factor of the GGPP pathway cascade such as I- κ B α [167, 168]. In the same way, statins induce the expression and secretion of tissue inhibitor of metalloproteinase 1 (TIMP-1) by SMCs and macrophages [124]. Regarding thrombosis, statin treatment has been shown *in vitro* and *in vivo* to reduce greatly the expression of tissue factor by vascular cells [104, 166]. Major determinants of the fibrolytic balance, such as plasminogen activator inhibitor-1 (PAI-1) and the tissue-type plasminogen activator (tPA) are also reduced by statin treatments. Indeed, simvastatin has been shown to reduce the secretion of PAI-1 from endothelial cells and SMCs, while the release of tPA was increased at the same concentrations [105]. This effect was linked to geranylgeranyl-modified intermediates, since it could be mimicked by C3 exoenzyme and prevented by GGPP, but not by FPP [105]. All these recent findings indicate that beyond their lipid-lowering effects, statins may reduce important inflammatory processes implicated in atherogenesis and plaque rupture that might in part also explain their great clinical benefits in cardiovascular diseases.

Statins as immunomodulators

Although a few clinical trials have suggested a better outcome of cardiac transplantation for patients on statin therapy [169, 170], statins have never been shown to be involved in the immune response. Major histocompatibility complex MHC class II (MHC-II) molecules, expressed on the surface of specialized cells, play a direct role in the control of the immune response and thus determine rejection after organ transplantation. Whereas a limited number of specialized cell-types express MHC-II constitutively, numerous other cells become MHC-II positive upon induction by the inflammatory mediator IFN- γ [171], including human endothelial cells and monocytes. This complex regulation is under the control of the transactivator CIITA [172, 173]. In recent studies, we showed that four different statins were able to inhibit inducible MHC-II expression in primary human endothelial cells and human monocyte/macrophages via inhibition of the promotor IV of the transactivator CIITA [174, 175]. These findings were confirmed by other reports [176, 177]. Since interactions between the endothelium and immunocompetent cells are crucial in the pathogenesis of cardiac allograft vasculopathy, our findings may represent one among other important mechanisms by which statins exert their beneficial effects after cardiac transplantation [169, 170, 178].

Therapy and prevention

The development of cholesterol-lowering agents helped to elucidate the implication of LDL cholesterol in cardiovascular diseases. Extensive clinical trials have demonstrated that some agents could reduce cardiovascular events, including MI, stroke, and death, by altering vascular atherosclerosis development in patients with or without previous symptoms of vascular artery disease [70, 134, 179–185]. Bile-sequestering resins, fibrates, and nicotinic acid (NA) have been used to reduce cholesterol, but were modestly effective and only poorly tolerated with low compliance [14]. Fibrates and NA have even inherent toxicities as they amplify the risk of hepatitis and myopathy. Indeed, several studies showed that these older lipid-altering agents increase the risk of non-cardiovascular death, and hence do not contribute to reducing total mortality.

Since their discovery in 1973, statins have been described as the principal and the most effective agents used to reduce serum cholesterol levels. Statins, or HMG CoA reductase inhibitors, have proven their beneficial effects in reducing total and LDL cholesterol as well as triglycerides, and increasing HDL cholesterol. Statin therapy has been shown by overwhelming clinical evidence to reduce significantly the risk of new or recurrent cardiovascular events and to improve survival of patients with pre-

vious cardiovascular disease [17]. In addition, statin treatments have demonstrated a marked reduction of ischemic strokes, the third leading cause of death in the United States after coronary disease and cancer. This effect on stroke was seen in patients whether or not they had suffered from previous vascular diseases (LIPID, CARE, WOSCOPS, 4S, HPS, and GREACE studies) [134, 144, 179, 180, 185–188]. In addition, evidence is increasing to highlight the importance of statin molecules with their beneficial effects on nonlipid mechanisms that control vascular immunoinflammatory processes.

Summary

For almost a decade, the use of statin therapy to treat dyslipidemia has demonstrated great beneficial effects, with reduction of cardiovascular morbidity and mortality in primary and secondary prevention. These large clinical benefits may not only be due to the lipid lowering effects of statins, but also to their pleiotropic effects. These new nonlipid vascular properties include: the improvement of endothelial function, anti-inflammation, anti-atherothrombosis, immunomodulation, antiproliferation, and antimigration. Together, these new findings of vascular effects of statins help us to better understand their great clinical benefits and may also provide a scientific rationale for their use in other diseases, such as rheumatoid arthritis, multiple sclerosis, Alzheimer's disease and following organ transplantation.

Acknowledgements. This work was supported by a grant from the Swiss National Science Foundation (3200-065121.01/1) to F. Mach and by the Foundation for Medical Research to N. Veillard.

- Navab M., Berliner J. A., Watson A. D., Hama S. Y., Territo M. C., Lusis A. J. et al. (1996) The Yin and Yang of oxidation in the development of the fatty streak: a review based on the 1994 George Lyman Duff Memorial Lecture. *Arterioscler. Thromb. Vasc. Biol.* **16**: 831–842
- Glass C. K. and Witztum J. L. (2001) Atherosclerosis: the road ahead. *Cell* **104**: 503–516
- Libby P., Ridker P. M. and Maseri A. (2002) Inflammation and atherosclerosis. *Circulation* **105**: 1135–1143
- Rosamond W. D., Chambless L. E., Folsom A. R., Cooper L. S., Conwill D. E., Clegg L. et al. (1998) Trends in the incidence of myocardial infarction and in mortality due to coronary heart disease, 1987 to 1994. *N. Engl. J. Med.* **339**: 861–867
- Heart and Stroke Statistical Update (2001) American Heart Association, Dallas, Tex.
- Post-Stroke Rehabilitation Guideline Panel No 16 (1995) Agency for Health Care Policy and Research, 95-0662
- Bryan R. N., Wells S. W., Miller T. J., Elster A. D., Jungreis C. A., Poirier V. C. et al. (1997) Infarct like lesions in the brain: prevalence and anatomic characteristics at MR imaging of the elderly – data from the Cardiovascular Health Study. *Radiology* **202**: 47–54
- Spieker L. E., Hürliemann D., Ruschitzka F., Corti R., Enseleit F., Shaw S. et al. (in press) Mental stress induces prolonged endothelial dysfunction via endothelin-A receptors. *Circulation* **105**
- Macleod J., Davey Smith G., Heslop P., Metcalfe C., Carroll D. and Hart C. (2002) Psychological stress and cardiovascular disease: empirical demonstration of bias in a prospective observational study of Scottish men. *BMJ* **324**: 1247
- Endo A. (1992) The discovery and development of HMG-CoA reductase inhibitors. *J. Lipid Res.* **33**: 1569–1582
- Kuroda M., Hazama-Shimada Y. and Endo A. (1977) Inhibition of sterol synthesis by citrinin in a cell-free system from rat liver and yeast. *Biochim. Biophys. Acta* **486**: 254–259
- Endo A., Tsujita Y., Kuroda M. and Tanzawa K. (1977) Inhibition of cholesterol synthesis in vitro and in vivo by ML-236A and ML-236B, competitive inhibitors of 3-hydroxy-3-methylglutaryl-coenzyme A reductase. *Eur. J. Biochem.* **77**: 31–36
- Page C. P., Curtis M. J., Sutter M. C., Walter M. J. A. H. and Hoffman B. B. (1997). In: *Integrated Pharmacology*. pp 267–270, Mosby (ed).
- Black D. M. (2002) A general assessment of the safety of HMG CoA reductase inhibitors (statins). *Curr. Atheroscler. Rep.* **4**: 34–41
- Corsini A., Bellosta S., Baetta R., Fumagalli R., Paoletti R. and Bernini F. (1999) New insights into the pharmacodynamic and pharmacokinetic properties of statins. *Pharmacol. Ther.* **84**: 413–428
- Serajuddin A. T., Ranadive S. A. and Mahoney E. M. (1991) Relative lipophilicities, solubilities, and structure-pharmacological considerations of 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase inhibitors pravastatin, lovastatin, mevastatin, and simvastatin. *J. Pharm. Sci.* **80**: 830–834
- LaRosa J. C., He J. and Vupputuri S. (1999) Effect of statins on risk of coronary disease: a meta-analysis of randomized controlled trials. *JAMA* **282**: 2340–2346
- Furberg C. D. (1999) Natural statins and stroke risk. *Circulation* **99**: 185–188
- Stein E., Plotkin D., Bays H., Davidson M., Dujovne C., Korenman S. et al. (2000) Effects of simvastatin (40 and 80 mg/day) in patients with mixed hyperlipidemia. *Am. J. Cardiol.* **86**: 406–411
- Jones P., Kafonek S., Laurora I. and Hunninghake D. (1998) Comparative dose efficacy study of atorvastatin versus simvastatin, pravastatin, lovastatin, and fluvastatin in patients with hypercholesterolemia (the CURVES study). *Am. J. Cardiol.* **81**: 582–587
- Goldstein J. L. and Brown M. S. (1990) Regulation of the mevalonate pathway. *Nature* **343**: 425–430
- Rosenson R. S. and Tangney C. C. (1998) Antiatherothrombotic properties of statins: implications for cardiovascular event reduction. *JAMA* **279**: 1643–1650
- Bernini F., Didoni G., Bonfadini G., Bellosta S. and Fumagalli R. (1993) Requirement for mevalonate in acetylated LDL induction of cholesterol esterification in macrophages. *Atherosclerosis* **104**: 19–26
- Bellosta S., Bernini F., Ferri N., Quarato P., Canavesi M., Arnaboldi L. et al. (1998) Direct vascular effects of HMG-CoA reductase inhibitors. *Atherosclerosis* **137** (suppl): S101–S109
- Glomset J. A., Gelb M. H. and Farnsworth C. C. (1990) Prenyl proteins in eukaryotic cells: a new type of membrane anchor. *Trends Biochem. Sci.* **15**: 139–142
- Maltese W. A. (1990) Posttranslational modification of proteins by isoprenoids in mammalian cells. *FASEB J.* **4**: 3319–3328
- Van Aelst L. and D'Souza-Schorey C. (1997) Rho GTPases and signaling networks. *Genes Dev.* **11**: 2295–2322
- Laufs U. and Liao J. K. (1998) Post-transcriptional regulation of endothelial nitric oxide synthase mRNA stability by Rho GTPase. *J. Biol. Chem.* **273**: 24266–24271

- 29 Laufs U., La Fata V., Plutzky J. and Liao J. K. (1998) Upregulation of endothelial nitric oxide synthase by HMG CoA reductase inhibitors. *Circulation* **97**: 1129–1135
- 30 Hall A. (1998) Rho GTPases and the actin cytoskeleton. *Science* **279**: 509–514
- 31 Laufs U. and Liao J. K. (2000) Targeting Rho in cardiovascular disease. *Circ. Res.* **87**: 526–528
- 32 Laufs U., Endres M., Stagliano N., Amin-Hanjani S., Chui D. S., Yang S. X. et al. (2000) Neuroprotection mediated by changes in the endothelial actin cytoskeleton. *J. Clin. Invest.* **106**: 15–24
- 33 Laufs U., Marra D., Node K. and Liao J. K. (1999) 3-Hydroxy-3-methylglutaryl-CoA reductase inhibitors attenuate vascular smooth muscle proliferation by preventing rho GTPase-induced down-regulation of p27 (Kip1). *J. Biol. Chem.* **274**: 21926–21931
- 34 Mundy G., Garrett R., Harris S., Chan J., Chen D., Rossini G. et al. (1999) Stimulation of bone formation in vitro and in rodents by statins. *Science* **286**: 1946–1949
- 35 Singh R., Wang B., Shirvaikar A., Khan S., Kamat S., Schelling J. R. et al. (1999) The IL-1 receptor and Rho directly associate to drive cell activation in inflammation. *J. Clin. Invest.* **103**: 1561–1570
- 36 Baetta R., Donetti E., Comparato C., Calore M., Rossi A., Teruzzi C. et al. (1997) Proapoptotic effect of atorvastatin on stimulated rabbit smooth muscle cells. *Pharmacol. Res.* **36**: 115–121
- 37 Essig M., Nguyen G., Prie D., Escoubet B., Sraer J. D. and Friedlander G. (1998) 3-Hydroxy-3-methylglutaryl coenzyme A reductase inhibitors increase fibrinolytic activity in rat aortic endothelial cells: role of geranylgeranylation and Rho proteins. *Circ. Res.* **83**: 683–690
- 38 Hernandez-Perera O., Perez-Sala D., Navarro-Antolin J., Sanchez-Pascuala R., Hernandez G., Diaz C. et al. (1998) Effects of the 3-hydroxy-3-methylglutaryl-CoA reductase inhibitors, atorvastatin and simvastatin, on the expression of endothelin-1 and endothelial nitric oxide synthase in vascular endothelial cells. *J. Clin. Invest.* **101**: 2711–2719
- 39 Munro E., Patel M., Chan P., Betteridge L., Clunn G., Gallagher K. et al. (1994) Inhibition of human vascular smooth muscle cell proliferation by lovastatin: the role of isoprenoid intermediates of cholesterol synthesis. *Eur. J. Clin. Invest.* **24**: 766–772
- 40 Martin G., Duez H., Blanquart C., Berezowski V., Poulain P., Fruchart J. C. et al. (2001) Statin-induced inhibition of the Rho-signaling pathway activates PPARalpha and induces HDL apoA-I. *J. Clin. Invest.* **107**: 1423–1432
- 41 Inoue I., Itoh F., Aoyagi S., Tazawa S., Kusama H., Akahane M. et al. (2002) Fibrate and statin synergistically increase the transcriptional activities of PPARalpha/RXRalpha and decrease the transactivation of NFkappaB. *Biochem. Biophys. Res. Commun.* **290**: 131–139
- 42 Mangelsdorf D. J., Thummel C., Beato M., Herrlich P., Schutz G., Umesono K. et al. (1995) The nuclear receptor superfamily: the second decade. *Cell* **83**: 835–839
- 43 Auboeuf D., Rieusset J., Fajas L., Vallier P., Frering V., Riou J. P. et al. (1997) Tissue distribution and quantification of the expression of mRNAs of peroxisome proliferator-activated receptors and liver X receptor-alpha in humans: no alteration in adipose tissue of obese and NIDDM patients. *Diabetes* **46**: 1319–1327
- 44 Granneman J., Skoff R. and Yang X. (1998) Member of the peroxisome proliferator-activated receptor family of transcription factors is differentially expressed by oligodendrocytes. *J. Neurosci. Res.* **51**: 563–573
- 45 Braissant O. and Wahli W. (1998) Differential expression of peroxisome proliferator-activated receptor-PPAR- α , - β and - γ during rat embryonic development. *Endocrinology* **139**: 2748–2754
- 46 Schoonjans K., Staels B. and Auwerx J. (1996) The peroxisome proliferator activated receptors (PPARS) and their effects on lipid metabolism and adipocyte differentiation. *Biochim. Biophys. Acta* **1302**: 93–109
- 47 Chinetti G., Fruchart J. C. and Staels B. (2001) Peroxisome proliferator-activated receptors (PPARs): nuclear receptors with functions in the vascular wall. *Z. Kardiol.* **90**: 125–132
- 48 Duez H., Fruchart J. C. and Staels B. (2001) PPARs in inflammation, atherosclerosis and thrombosis. *J. Cardiovasc. Risk* **8**: 187–194
- 49 Marx N., Kehrle B., Kohlhammer K., Grub M., Koenig W., Hombach V. et al. (2002) PPAR activators as antiinflammatory mediators in human T lymphocytes: implications for atherosclerosis and transplantation-associated arteriosclerosis. *Circ. Res.* **90**: 703–710
- 50 Ricote M., Li A. C., Willson T. M., Kelly C. J. and Glass C. K. (1998) The peroxisome proliferator-activated receptor-gamma is a negative regulator of macrophage activation. *Nature* **391**: 79–82
- 51 Staels B., Koenig W., Habib A., Merval R., Lebret M., Torra I. P. et al. (1998) Activation of human aortic smooth-muscle cells is inhibited by PPARalpha but not by PPARgamma activators. *Nature* **393**: 790–793
- 52 Delerive P., De Bosscher K., Besnard S., Vanden Berghe W., Peters J. M., Gonzalez F. J. et al. (1999) Peroxisome proliferator-activated receptor alpha negatively regulates the vascular inflammatory gene response by negative cross-talk with transcription factors NF-kappaB and AP-1. *J. Biol. Chem.* **274**: 32048–32054
- 53 Brand K., Page S., Rogler G., Bartsch A., Brandl R., Knuechel R. et al. (1996) Activated transcription factor nuclear factor-kappa B is present in the atherosclerotic lesion. *J. Clin. Invest.* **97**: 1715–1722
- 54 Bustos C., Hernandez-Presa M. A., Ortego M., Tunon J., Ortega L., Perez F. et al. (1998) HMG-CoA reductase inhibition by atorvastatin reduces neointimal inflammation in a rabbit model of atherosclerosis. *J. Am. Coll. Cardiol.* **32**: 2057–2064
- 55 Mach F. (2001) The role of chemokines in atherosclerosis. *Curr. Atheroscler. Rep.* **3**: 243–251
- 56 Bourcier T., Sukhova G. and Libby P. (1997) The nuclear factor kappa-B signaling pathway participates in dysregulation of vascular smooth muscle cells in vitro and in human atherosclerosis. *J. Biol. Chem.* **272**: 15817–15824
- 57 Ueda A., Okuda K., Ohno S., Shirai A., Igarashi T., Matsunaga K. et al. (1994) NF-kappa B and Sp1 regulate transcription of the human monocyte chemoattractant protein-1 gene. *J. Immunol.* **153**: 2052–2063
- 58 Dain J. G., Fu E., Gorski J., Nicoletti J. and Scallen T. J. (1993) Biotransformation of fluvastatin sodium in humans. *Drug Metab. Dispos.* **21**: 567–572
- 59 Christians U., Jacobsen W. and Floren L. C. (1998) Metabolism and drug interactions of 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors in transplant patients: are the statins mechanistically similar? *Pharmacol. Ther.* **80**: 1–34
- 60 Quion J. A. and Jones P. H. (1994) Clinical pharmacokinetics of pravastatin. *Clin. Pharmacokinet.* **27**: 94–103
- 61 Singhvi S. M., Pan H. Y., Morrison R. A. and Willard D. A. (1990) Disposition of pravastatin sodium, a tissue-selective HMG-CoA reductase inhibitor, in healthy subjects. *Br. J. Clin. Pharmacol.* **29**: 239–243
- 62 McClellan K. J., Wiseman L. R. and McTavish D. (1998) Cerivastatin. *Drugs* **55**: 415–420
- 63 Lennernas H. and Fager G. (1997) Pharmacodynamics and pharmacokinetics of the HMG-CoA reductase inhibitors: similarities and differences. *Clin. Pharmacokinet.* **32**: 403–425
- 64 Becquemont L. (2000) Inhibiteurs de l'HMG-CoA réductase et interactions médicamenteuses. *Lettre Pharmacol.* **14**: 64–68

- 65 Fischer V., Johanson L., Heitz F., Tullman R., Graham E., Baldeck J. P. et al. (1999) The 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitor fluvastatin: effect on human cytochrome P-450 and implications for metabolic drug interactions. *Drug Metab. Dispos.* **27**: 410–416
- 66 Hodel C. (2002) Myopathy and rhabdomyolysis with lipid-lowering drugs. *Toxicol. Lett.* **128**: 159–168
- 67 Staffa J. A., Chang J. and Green L. (2002) Cerivastatin and reports of fatal rhabdomyolysis. *N. Engl. J. Med.* **346**: 539–540
- 68 McQueen M. J. (1990) Cholestatic jaundice associated with lovastatin (Mevacor) therapy. *Can. Med. Assoc. J.* **142**: 841–842
- 69 Theal R. M. and Scott K. (1996) Evaluating asymptomatic patients with abnormal liver function test results. *Am. Fam. Physician* **53**: 2111–2119
- 70 Downs J. R., Clearfield M., Weis S., Whitney E., Shapiro D. R., Beere P. A. et al. (1998) Primary prevention of acute coronary events with lovastatin in men and women with average cholesterol levels: results of AFCAPS/TexCAPS. Air Force/Texas Coronary Atherosclerosis Prevention Study. *JAMA* **279**: 1615–1622
- 71 Pekkanen J., Linn S., Heiss G., Suchindran C. M., Leon A., Rifkind B. M. et al. (1990) Ten-year mortality from cardiovascular disease in relation to cholesterol level among men with and without preexisting cardiovascular disease. *N. Engl. J. Med.* **322**: 1700–1707
- 72 Pekkanen J., Tervahauta M., Nissinen A. and Karvonen M. J. (1993) Does the predictive value of baseline coronary risk factors change over a 30-year follow-up? *Cardiology* **82**: 181–190
- 73 The Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (1993) Summary of the second report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel II). *JAMA* **269**: 3015–3023
- 74 Grover S. A., Coupal L. and Hu X. P. (1995) Identifying adults at increased risk of coronary disease: how well do the current cholesterol guidelines work? *JAMA* **274**: 801–806
- 75 Castelli W. P., Garrison R. J., Wilson P. W., Abbott R. D., Kalousdian S. and Kannel W. B. (1986) Incidence of coronary heart disease and lipoprotein cholesterol levels. The Framingham Study *JAMA* **256**: 2835–2838
- 76 Thompson S. G., Kienast J., Pyke S. D., Haverkate F. and Loo J. C. van de (1995) Hemostatic factors and the risk of myocardial infarction or sudden death in patients with angina pectoris. European Concerted Action on Thrombosis and Disabilities Angina Pectoris Study Group. *N. Engl. J. Med.* **332**: 635–641
- 77 Heinrich J., Balleisen L., Schulte H., Assmann G. and Loo J. van de (1994) Fibrinogen and factor VII in the prediction of coronary risk: results from the PROCAM study in healthy men. *Arterioscler. Thromb.* **14**: 54–59
- 78 Sweetnam P. M., Thomas H. F., Yarnell J. W., Beswick A. D., Baker I. A. and Elwood P. C. (1996) Fibrinogen, viscosity and the 10-year incidence of ischaemic heart disease. *Eur. Heart. J.* **17**: 1814–1820
- 79 Scott H. D. and Laake K. (2001) Statins for the prevention of Alzheimer's disease. *Cochrane Database Syst. Rev.* **4**
- 80 Fassbender K., Simons M., Bergmann C., Stroick M., Lutjohann D., Keller P. et al. (2001) Simvastatin strongly reduces levels of Alzheimer's disease beta-amyloid peptides Abeta 42 and Abeta 40 in vitro and in vivo. *Proc. Natl. Acad. Sci.* **98**: 5856–5861
- 81 Corder E. H., Saunders A. M., Strittmatter W. J., Schmechel D. E., Gaskell P. C., Small G. W. et al. (1993) Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* **261**: 921–923
- 82 Hofman A., Ott A., Breteler M. M., Bots M. L., Slooter A. J., van Harskamp F. van et al. (1997) Atherosclerosis, apolipoprotein E, and prevalence of dementia and Alzheimer's disease in the Rotterdam Study. *Lancet* **349**: 151–154
- 83 Bales K. R., Verina T., Dodel R. C., Du Y., Altstiel L., Bender M. et al. (1997) Lack of apolipoprotein E dramatically reduces amyloid beta-peptide deposition. *Nat. Genet.* **17**: 263–264
- 84 Refolo L. M., Malester B., LaFrancois J., Bryant-Thomas T., Wang R., Tint G. S. et al. (2000) Hypercholesterolemia accelerates the Alzheimer's amyloid pathology in a transgenic mouse model. *Neurobiol. Dis.* **7**: 321–331
- 85 Sparks D. L., Scheff S. W., Hunsaker J. C., 3rd Liu H., Landers T. and Gross D. R. (1994) Induction of Alzheimer-like beta-amyloid immunoreactivity in the brains of rabbits with dietary cholesterol. *Exp. Neurol.* **126**: 88–94
- 86 Simons M., Keller P., De Strooper B., Beyreuther K., Dotti C. G. and Simons K. (1998) Cholesterol depletion inhibits the generation of beta-amyloid in hippocampal neurons. *Proc. Natl. Acad. Sci. USA* **95**: 6460–6464
- 87 Wolozin B., Kellman W., Ruosseau P., Celesia G. G. and Siegel G. (2000) Decreased prevalence of Alzheimer disease associated with 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors. *Arch. Neurol.* **57**: 1439–1443
- 88 Mach F., Sauty A., Iarossi A. S., Sukhova G. K., Neote K., Libby P. et al. (1999) Differential expression of three T lymphocyte-activating CXC chemokines by human atheroma-associated cells. *J. Clin. Invest.* **104**: 1041–1050
- 89 Ignarro L. J. (1990) Biosynthesis and metabolism of endothelium-derived nitric oxide. *Annu. Rev. Pharmacol. Toxicol.* **30**: 535–560
- 90 Palmer R. M., Ferrige A. G. and Moncada S. (1987) Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor. *Nature* **327**: 524–526
- 91 Radomski M. W., Palmer R. M. and Moncada S. (1990) An L-arginine/nitric oxide pathway present in human platelets regulates aggregation. *Proc. Natl. Acad. Sci. USA* **87**: 5193–5197
- 92 Huang P. L., Huang Z., Mashimo H., Bloch K. D., Moskowitz M. A., Bevan J. A. et al. (1995) Hypertension in mice lacking the gene for endothelial nitric oxide synthase. *Nature* **377**: 239–242
- 93 Furchgott R. F. and Zawadzki J. V. (1980) The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature* **288**: 373–376
- 94 Huang Z., Huang P. L., Ma J., Meng W., Ayata C., Fishman M. C. et al. (1996) Enlarged infarcts in endothelial nitric oxide synthase knockout mice are attenuated by nitro-L-arginine. *J. Cereb. Blood Flow Metab.* **16**: 981–987
- 95 Huang Z., Huang P. L., Panahian N., Dalkara T., Fishman M. C. and Moskowitz M. A. (1994) Effects of cerebral ischemia in mice deficient in neuronal nitric oxide synthase. *Science* **265**: 1883–1885
- 96 Morikawa E., Huang Z. and Moskowitz M. A. (1992) L-arginine decreases infarct size caused by middle cerebral arterial occlusion in SHR. *Am. J. Physiol.* **263**: H1632–H1635
- 97 Zhang F. and Iadecola C. (1994) Reduction of focal cerebral ischemic damage by delayed treatment with nitric oxide donors. *J. Cereb. Blood Flow Metab.* **14**: 574–580
- 98 Dalkara T., Morikawa E., Panahian N. and Moskowitz M. A. (1994) Blood flow-dependent functional recovery in a rat model of focal cerebral ischemia. *Am. J. Physiol.* **267**: H678–H683
- 99 Laufs U., Fata V. L. and Liao J. K. (1997) Inhibition of 3-hydroxy-3-methylglutaryl (HMG)-CoA reductase blocks hypoxia-mediated down-regulation of endothelial nitric oxide synthase. *J. Biol. Chem.* **272**: 31725–31729
- 100 Liao J. K., Zulueta J. J., Yu F. S., Peng H. B., Cote C. G. and Hassoun P. M. (1995) Regulation of bovine endothelial constitutive nitric oxide synthase by oxygen. *J. Clin. Invest.* **96**: 2661–2666
- 101 Endres M., Laufs U., Huang Z., Nakamura T., Huang P., Moskowitz M. A. et al. (1998) Stroke protection by 3-hy-

- droxy-3-methylglutaryl (HMG)-CoA reductase inhibitors mediated by endothelial nitric oxide synthase. *Proc. Natl. Acad. Sci. USA* **95**: 8880–8885
- 102 Stalker T. J., Lefer A. M. and Scalia R. (2001) A new HMG-CoA reductase inhibitor, rosuvastatin, exerts anti-inflammatory effects on the microvascular endothelium: the role of mevalonic acid. *Br. J. Pharmacol.* **133**: 406–412
 - 103 Rikitake Y., Kawashima S., Takeshita S., Yamashita T. et al. (2001) Anti-oxidative properties of fluvastatin, an HMG-CoA reductase inhibitor, contribute to prevention of atherosclerosis in cholesterol-fed rabbits. *Atherosclerosis* **154**: 87–96
 - 104 Eto M., Kozai T., Cosentino F., Joch H. and Lüscher T. F. (2002) Statin prevents tissue factor expression in human endothelial cells: role of Rho/Rho-kinase and Akt pathways. *Circulation* **105**: 1756–1759
 - 105 Bourcier T. and Libby P. (2000) HMG CoA reductase inhibitors reduce plasminogen activator inhibitor-1 expression by human vascular smooth muscle and endothelial cells. *Arterioscler. Thromb. Vasc. Biol.* **20**: 556–562
 - 106 Levin E. R. (1995) Endothelins. *N. Engl. J. Med.* **333**: 356–363
 - 107 Heeschen C., Hamm C.W., Laufs U., Snapinn S., Böhm M. and White H.D. (2002) Withdrawal of statins increases event rates in patients with acute coronary syndromes. *Circulation* **105**: 1446–1452
 - 108 Moreno P. R., Falk E., Palacios I. F., Newell J. B., Fuster V. and Fallon J. T. (1994) Macrophage infiltration in acute coronary syndromes: implications for plaque rupture. *Circulation* **90**: 775–778
 - 109 Romano M., Mezzetti A., Marulli C., Ciabattini G., Febo F., Di Ienno S. et al. (2000) Fluvastatin reduces soluble P-selectin and ICAM-1 levels in hypercholesterolemic patients: role of nitric oxide. *J. Invest. Med.* **48**: 183–189
 - 110 Nakashima Y., Raines E. W., Plump A. S., Breslow J. L. and Ross R. (1998) Upregulation of VCAM-1 and ICAM-1 at atherosclerosis-prone sites on the endothelium in the ApoE-deficient mouse. *Arterioscler. Thromb. Vasc. Biol.* **18**: 842–851
 - 111 Shih P. T., Brennan M. L., Vora D. K., Territo M. C., Strahl D., Elices M. J. et al. (1999) Blocking very late antigen-4 integrin decreases leukocyte entry and fatty streak formation in mice fed an atherogenic diet. *Circ. Res.* **84**: 345–351
 - 112 Nie Q., Fan J., Haraoka S., Shimokama T. and Watanabe T. (1997) Inhibition of mononuclear cell recruitment in aortic intima by treatment with anti-ICAM-1 and anti-LFA-1 monoclonal antibodies in hypercholesterolemic rats: implications of the ICAM-1 and LFA-1 pathway in atherogenesis. *Lab. Invest.* **77**: 469–482
 - 113 Weber C., Erl W., Weber K. S. and Weber P. C. (1997) HMG-CoA reductase inhibitors decrease CD11b expression and CD11b-dependent adhesion of monocytes to endothelium and reduce increased adhesiveness of monocytes isolated from patients with hypercholesterolemia. *J. Am. Coll. Cardiol.* **30**: 1212–1217
 - 114 Weitz-Schmidt G., Welzenbach K., Brinkmann V., Kamata T., Kallen J., Bruns C. et al. (2001) Statins selectively inhibit leukocyte function antigen-1 by binding to a novel regulatory integrin site. *Nat. Med.* **7**: 687–692
 - 115 Rasmussen L. M., Hansen P. R., Nabipour M. T., Olesen P., Kristiansen M. T. and Ledet T. (2001) Diverse effects of inhibition of 3-hydroxy-3-methylglutaryl-CoA reductase on the expression of VCAM-1 and E-selectin in endothelial cells. *Biochem. J.* **360**: 363–370
 - 116 Romano M., Diomedea L., Sironi M., Massimiliano L., Sottocorno M., Polentarutti N. et al. (2000) Inhibition of monocyte chemotactic protein-1 synthesis by statins. *Lab. Invest.* **80**: 1095–1100
 - 117 Pahan K., Sheikh F. G., Nambodiri A. M. and Singh I. (1997) Lovastatin and phenylacetate inhibit the induction of nitric oxide synthase and cytokines in rat primary astrocytes, microglia, and macrophages. *J. Clin. Invest.* **100**: 2671–2679
 - 118 Kothe H., Dalhoff K., Rupp J., Müller A., Kreuzer J., Maass M. et al. (2000) Hydroxymethylglutaryl coenzyme A reductase inhibitors modify the inflammatory response of human macrophages and endothelial cells infected with *Chlamydia pneumoniae*. *Circulation* **101**: 1760–1763
 - 119 Ikeda U. and Shimada K. (1999) Statins and monocytes. *Lancet* **353**: 2070
 - 120 Ikeda U., Hojo Y., Katsuki T. and Shimada K. (1999) Procoagulant and proinflammatory activity in acute coronary syndromes. *Cardiovasc. Res.* **42**: 823–824
 - 121 Rudich S. M., Mongini P. K., Perez R. V. and Katznelson S. (1998) HMG-CoA reductase inhibitors pravastatin and simvastatin inhibit human B-lymphocyte activation. *Transplant. Proc.* **30**: 992–995
 - 122 Garlicks C. D., John S., Schmeisser A., Eskafi S., Stumpf C., Karl M. et al. (2001) Upregulation of CD40 and CD40 ligand (CD154) in patients with moderate hypercholesterolemia. *Circulation* **104**: 2395–2400
 - 123 Vaughan C. J., Gotto A. M. Jr and Basson C. T. (2000) The evolving role of statins in the management of atherosclerosis. *J. Am. Coll. Cardiol.* **35**: 1–10
 - 124 Crisby M., Nordin-Fredriksson G., Shah P. K., Yano J., Zhu J. and Nilsson J. (2001) Pravastatin treatment increases collagen content and decreases lipid content, inflammation, metalloproteinases, and cell death in human carotid plaques: implications for plaque stabilization. *Circulation* **103**: 926–933
 - 125 Yoshida M., Sawada T., Ishii H., Gerszten R.E., Rosenzweig A., Gimbrone M. A. Jr et al. (2001) HMG-CoA reductase inhibitor modulates monocyte-endothelial cell interaction under physiological flow conditions in vitro: involvement of Rho GTPase-dependent mechanism. *Arterioscler. Thromb. Vasc. Biol.* **21**: 1165–1171
 - 126 Wojciak-Stothard B., Williams L. and Ridley A. J. (1999) Monocyte adhesion and spreading on human endothelial cells is dependent on Rho-regulated receptor clustering. *J. Cell. Biol.* **145**: 1293–1307
 - 127 Liu L., Moesner P., Kovach N. L., Bailey R., Hamilton A. D., Sebt S. M. et al. (1999) Integrin-dependent leukocyte adhesion involves geranylgeranylated protein(s). *J. Biol. Chem.* **274**: 33334–33340
 - 128 Patel K. D., Zimmerman G. A., Prescott S. M., McEver R. P. and McIntyre T. M. (1991) Oxygen radicals induce human endothelial cells to express GMP-140 and bind neutrophils. *J. Cell. Biol.* **112**: 749–759
 - 129 Davenpeck K. L., Gauthier T. W. and Lefer A. M. (1994) Inhibition of endothelial-derived nitric oxide promotes P-selectin expression and actions in the rat microcirculation. *Gastroenterology* **107**: 1050–1058
 - 130 Baumann H. and Gauldie J. (1994) The acute phase response. *Immunol. Today* **15**: 74–80
 - 131 Ridker P. M., Rifai N., Clearfield M., Downs J. R., Weis S. E., Miles J. S. et al. (2001) Measurement of C-reactive protein for the targeting of statin therapy in the primary prevention of acute coronary events. *N. Engl. J. Med.* **344**: 1959–1965
 - 132 Liuzzo G., Biasucci L. M., Gallimore J. R., Grillo R. L., Rebuzzi A. G., Pepys M. B. et al. (1994) The prognostic value of C-reactive protein and serum amyloid A protein in severe unstable angina. *N. Engl. J. Med.* **331**: 417–424
 - 133 Ridker P. M., Hennekens C. H., Buring J. E. and Rifai N. (2000) C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women. *N. Engl. J. Med.* **342**: 836–843
 - 134 Sacks F. M., Pfeffer M. A., Moye L. A., Rouleau J. L., Rutherford J. D., Cole T. G. et al. (1996) The effect of pravastatin on coronary events after myocardial infarction in patients with average cholesterol levels. Cholesterol and Recurrent Events Trial investigators. *N. Engl. J. Med.* **335**: 1001–1009

- 135 Ridker P. M., Rifai N. and Lowenthal S. P. (2001) Rapid reduction in C-reactive protein with cerivastatin among 785 patients with primary hypercholesterolemia. *Circulation* **103**: 1191–1193
- 136 Musial J., Undas A., Gajewski P., Jankowski M., Sydor W. and Szczeklik A. (2001) Anti-inflammatory effects of simvastatin in subjects with hypercholesterolemia. *Int. J. Cardiol.* **77**: 247–253
- 137 Ridker P. M., Rifai N., Pfeffer M. A., Sacks F. M., Moye L. A., Goldman S., Flaker G. C. et al. (1998) Inflammation, pravastatin, and the risk of coronary events after myocardial infarction in patients with average cholesterol levels. Cholesterol and Recurrent Events (CARE) Investigators. *Circulation*. **98**: 839–844
- 138 Wang X., Li X., Yue T. L. and Ohlstein E. H. (2000) Expression of monocyte chemotactic protein-3 mRNA in rat vascular smooth muscle cells and in carotid artery after balloon angioplasty. *Biochim. Biophys. Acta* **1500**: 41–48
- 139 Hayes I. M., Jordan N. J., Towers S., Smith G., Paterson J. R., Earnshaw J. J., Roach A. G., Westwick J. et al. (1998) Human vascular smooth muscle cells express receptors for CC chemokines. *Arterioscler. Thromb. Vasc. Biol.* **18**: 397–403
- 140 Raiteri M., Arnaboldi L., McGeady P., Gelb M. H., Verri D., Tagliabue C. et al. (1997) Pharmacological control of the mevalonate pathway: effect on arterial smooth muscle cell proliferation. *J. Pharmacol. Exp. Ther.* **281**: 1144–1153
- 141 Hidaka Y., Eda T., Yonemoto M. and Kamei T. (1992) Inhibition of cultured vascular smooth muscle cell migration by simvastatin (MK-733). *Atherosclerosis* **95**: 87–94
- 142 Mulder H. J., Bal E. T., Jukema J. W., Zwinderman A. H., Schalij M. J., Boven A. J. van et al. (2000) Pravastatin reduces restenosis two years after percutaneous transluminal coronary angioplasty (REGRESS trial). *Am. J. Cardiol.* **86**: 742–746
- 143 Walter D. H., Schachinger V., Elsner M., Mach S., Auch-Schwelk W. and Zeiher A. M. (2000) Effect of statin therapy on restenosis after coronary stent implantation. *Am. J. Cardiol.* **85**: 962–968
- 144 Serruys P. W., de Feyter P. de, Macaya C., Kokott N., Puel J., Vrolix M. et al. (2002) Fluvastatin for prevention of cardiac events following successful first percutaneous coronary intervention: a randomized controlled trial. *JAMA* **287**: 3215–322
- 145 Weinberg R. A. (1995) The retinoblastoma protein and cell cycle control. *Cell* **81**: 323–330
- 146 Hirai A., Nakamura S., Noguchi Y., Yasuda T., Kitagawa M., Tatsuno I. et al. (1997) Geranylgeranylated rho small GTPase(s) are essential for the degradation of p27Kip1 and facilitate the progression from G1 to S phase in growth-stimulated rat FRTL-5 cells. *J. Biol. Chem.* **272**: 13–16
- 147 Rogler G., Lackner K. J. and Schmitz G. (1995) Effects of fluvastatin on growth of porcine and human vascular smooth muscle cells in vitro. *Am. J. Cardiol.* **76**: 114A–116A
- 148 Soma M. R., Donetti E., Parolini C., Mazzini G., Ferrari C., Fumagalli R. et al. (1993) HMG CoA reductase inhibitors: in vivo effects on carotid intimal thickening in normocholesterolemic rabbits. *Arterioscler. Thromb.* **13**: 571–578
- 149 Stark W. W. Jr, Blaskovich M. A., Johnson B. A., Qian Y., Vasudevan A., Pitt B. et al. (1998) Inhibiting geranylgeranylation blocks growth and promotes apoptosis in pulmonary vascular smooth muscle cells. *Am. J. Physiol.* **275**: L55–L63
- 150 Rembold C. (1996) Could atherosclerosis originate from defective smooth muscle cell death (apoptosis)? *Perspect. Biol. Med.* **39**: 405–408
- 151 Bochaton-Piallat M. L., Gabbiani F., Redard M., Desmouliere A. and Gabbiani G. (1995) Apoptosis participates in cellular regulation during rat aortic intimal thickening. *Am. J. Pathol.* **146**: 1059–1064
- 152 Guijarro C., Blanco-Colio L. M., Ortego M., Alonso C., Ortiz A., Plaza J. J. et al. (1998) 3-Hydroxy-3-methylglutaryl coenzyme A reductase and isoprenylation inhibitors induce apoptosis of vascular smooth muscle cells in culture. *Circ. Res.* **83**: 490–500
- 153 Li X., Liu L., Tupper J. C., Bannerman D. D., Winn R. K., Sebt S. M. et al. (2002) Inhibition of protein geranylgeranylation and RhoA/RhoA kinase pathway induces apoptosis in human endothelial cells. *J. Biol. Chem.* **277**: 15309–15316
- 154 Geng Y. J., Henderson L. E., Levesque E. B., Muszynski M. and Libby P. (1997) Fas is expressed in human atherosclerotic intima and promotes apoptosis of cytokine-primed human vascular smooth muscle cells. *Arterioscler. Thromb. Vasc. Biol.* **17**: 2200–2208
- 155 Maciejewski J., Selleri C., Anderson S. and Young N. S. (1995) Fas antigen expression on CD34+ human marrow cells is induced by interferon gamma and tumor necrosis factor alpha and potentiates cytokine-mediated hematopoietic suppression in vitro. *Blood* **85**: 3183–3190
- 156 Martinez-Gonzalez J., Vinals M., Vidal F., Llorente-Cortes V. and Badimon L. (1997) Mevalonate deprivation impairs IGF-I/insulin signaling in human vascular smooth muscle cells. *Atherosclerosis* **135**: 213–223
- 157 Ross R. (1993) The pathogenesis of atherosclerosis: a perspective for the 1990s. *Nature* **362**: 801–809
- 158 Davies M. J., Richardson P. D., Woolf N., Katz D. R. and Mann J. (1993) Risk of thrombosis in human atherosclerotic plaques: role of extracellular lipid, macrophage, and smooth muscle cell content. *Br. Heart J.* **69**: 377–381
- 159 Burleigh M. C., Briggs A. D., Lendon C. L., Davies M. J., Born G. V. and Richardson P. D. (1992) Collagen types I and III, collagen content, GAGs and mechanical strength of human atherosclerotic plaque caps: span-wise variations. *Atherosclerosis* **96**: 71–81
- 160 Bennett M. R. (1999) Apoptosis of vascular smooth muscle cells in vascular remodelling and atherosclerotic plaque rupture. *Cardiovasc. Res.* **41**: 361–368
- 161 Fuster V., Stein B., Ambrose J. A., Badimon L., Badimon J. J. and Chesebro J. H. (1990) Atherosclerotic plaque rupture and thrombosis: evolving concepts. *Circulation* **82**: II47–II59
- 162 Takemoto M. and Liao J. K. (2001) Pleiotropic effects of 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors. *Arterioscler. Thromb. Vasc. Biol.* **21**: 1712–1719
- 163 Aikawa M., Rabkin E., Voglic S. J., Shing H., Nagai R., Schoen F. J. et al. (1998) Lipid lowering promotes accumulation of mature smooth muscle cells expressing smooth muscle myosin heavy chain isoforms in rabbit atheroma. *Circ. Res.* **83**: 1015–1026
- 164 Bellosta S., Via D., Canavesi M., Pfister P., Fumagalli R., Paoletti R. et al. (1998) HMG-CoA reductase inhibitors reduce MMP-9 secretion by macrophages. *Arterioscler. Thromb. Vasc. Biol.* **18**: 1671–1678
- 165 Fukumoto Y., Libby P., Rabkin E., Hill C. C., Enomoto M., Hirouchi Y. et al. (2001) Statins alter smooth muscle cell accumulation and collagen content in established atheroma of watanabe heritable hyperlipidemic rabbits. *Circulation* **103**: 993–999
- 166 Aikawa M., Rabkin E., Sugiyama S., Voglic S. J., Fukumoto Y., Furukawa Y. et al. (2001) An HMG-CoA reductase inhibitor, cerivastatin, suppresses growth of macrophages expressing matrix metalloproteinases and tissue factor in vivo and in vitro. *Circulation* **103**: 276–283
- 167 Wong B., Lumma W. C., Smith A. M., Sisko J. T., Wright S. D. and Cai T. Q. (2001) Statins suppress THP-1 cell migration and secretion of matrix metalloproteinase 9 by inhibiting geranylgeranylation. *J. Leukoc. Biol.* **69**: 959–962
- 168 Chase A. J., Bond M., Crook M. F. and Newby A. C. (2002) Role of nuclear factor-kappaB activation in metalloproteinase-1, -3, and -9 secretion by human macrophages in vitro and rabbit foam cells produced in vivo. *Arterioscler. Thromb. Vasc. Biol.* **22**: 765–771
- 169 Kobashigawa J. A., Katznelson S., Laks H., Johnson J. A., Yeatman L., Wang X. M. et al. (1995) Effect of pravastatin on

- outcomes after cardiac transplantation. *N. Engl. J. Med.* **333**: 621–627
- 170 Wenke K., Meiser B., Thiery J., Nagel D., Scheidt W. von, Steinbeck G. et al. (1997) Simvastatin reduces graft vessel disease and mortality after heart transplantation: a four-year randomized trial. *Circulation* **96**: 1398–1402
 - 171 Reith W. and Mach B. (2001) The bare lymphocyte syndrome and the regulation of MHC expression. *Annu. Rev. Immunol.* **19**: 331–373
 - 172 Steimle V., Otten L. A., Zufferey M. and Mach B. (1993) Complementation cloning of an MHC class II transactivator mutated in hereditary MHC class II deficiency (or bare lymphocyte syndrome). *Cell* **75**: 135–146
 - 173 Steimle V., Siegrist C. A., Mottet A., Lisowska-Grospierre B. and Mach B. (1994) Regulation of MHC class II expression by interferon-gamma mediated by the transactivator gene CIITA. *Science* **265**: 106–109
 - 174 Kwak B., Mulhaupt F., Veillard N., Pelli G. and Mach F. (2001) The HMG-CoA reductase inhibitor simvastatin inhibits IFN-gamma induced MHC class II expression in human vascular endothelial cells. *Swiss Med. Wkly* **131**: 41–46
 - 175 Kwak B., Mulhaupt F., Myit S. and Mach F. (2000) Statins as a newly recognized type of immunomodulator. *Nat. Med.* **6**: 1399–1402
 - 176 Kwak B. R., Myit S., Mulhaupt F., Veillard N., Rufer N., Roosenek E. et al. (2002) PPARgamma but not PPARalpha ligands are potent repressors of major histocompatibility complex class II induction in atheroma-associated cells. *Circ. Res.* **90**: 356–362
 - 177 Sadeghi M. M., Tiglio A., Sadigh K., O'Donnell L., Collinge M., Pardi R. et al. (2001) Inhibition of interferon-gamma-mediated microvascular endothelial cell major histocompatibility complex class II gene activation by HMG-CoA reductase inhibitors. *Transplantation* **71**: 1262–1268
 - 178 Kobashigawa J. A. (2001) Statins as immunosuppressive agents. *Liver Transpl.* **7**: 559–561
 - 179 The Long-Term Intervention with Pravastatin in Ischaemic Disease (LIPID) Study Group (1998) Prevention of cardiovascular events and death with pravastatin in patients with coronary heart disease and a broad range of initial cholesterol levels. *N. Engl. J. Med.* **339**: 1349–1357
 - 180 The Scandinavian Simvastatin Survival Study (4S) (1994) Randomised trial of cholesterol lowering in 4444 patients with coronary heart disease. *Lancet* **344**: 1383–1389
 - 181 Jukema J. W., Bruschke A. V., Boven A. J. van, Reiber J. H., Bal E. T., Zwinderman A. H. et al. (1995) Effects of lipid lowering by pravastatin on progression and regression of coronary artery disease in symptomatic men with normal to moderately elevated serum cholesterol levels. The Regression Growth Evaluation Statin Study (REGRESS). *Circulation* **91**: 2528–2540
 - 182 Herd J. A., Ballantyne C. M., Farmer J. A., Ferguson J. J. 3rd, Jones P. H., West M. S. et al. (1997) Effects of fluvastatin on coronary atherosclerosis in patients with mild to moderate cholesterol elevations (Lipoprotein and Coronary Atherosclerosis Study [LCAS]). *Am. J. Cardiol.* **80**: 278–286
 - 183 Brown B. G., Zhao X. Q., Sacco D. E. and Albers J. J. (1993) Lipid lowering and plaque regression: new insights into prevention of plaque disruption and clinical events in coronary disease. *Circulation* **87**: 1781–1791
 - 184 Riegger G., Abletshauser C., Ludwig M., Schwandt P., Widimsky J., Weidinger G. et al. (1999) The effect of fluvastatin on cardiac events in patients with symptomatic coronary artery disease during one year of treatment. *Atherosclerosis* **144**: 263–270
 - 185 Shepherd J. (1995) Fibrates and statins in the treatment of hyperlipidaemia: an appraisal of their efficacy and safety. *Eur. Heart. J.* **16**: 5–13
 - 186 Heart Protection Study Collaborative Group. (2002) MRC/BHF Heart Protection Study of cholesterol lowering with simvastatin in 20 536 high-risk individuals: a randomized placebo-controlled trial. *Lancet.* **360**: 7–22
 - 187 Lescol Intervention Prevention Study (LIPS) investigators. (2002) Fluvastatin for prevention of cardiac events following successful first percutaneous coronary intervention. *JAMA.* **287**: 3215–3222
 - 188 Athyros V. G., Papageorgiou A. A., Mercouris B. R., Athyrou V. V., Symeonidis A. N., Basayannis E. O. and Demetriadis D. S. (2002) Treatment with atorvastatin to the national cholesterol educational program goal versus 'usual' care in secondary coronary heart disease prevention. The GREek Atorvastatin and Coronary-heart-disease Evaluation (GREACE) Study. *Curr. Med. Res. Opin.* **18**: 220–228



To access this journal online:
<http://www.birkhauser.ch>